

## NEUTRALIZATION AND BIOPRINTING PROTOCOL

# Coll 1 Solution

This is a suggested procedure, please adjust according to your experimental needs. To maintain the sterility of the product, work under sterile conditions.

### Protocol aim

The aim of this protocol is to provide instructions for neutralization and subsequent bioprinting of the neutralized Coll 1 Solution. This document covers the bioprinting of Coll 1 droplets with embedded cells and cell post-seeding on printed Coll 1 constructs. The biomaterial thermally gels at 37°C.

### Materials needed

- Coll 1 Solution (10 mL at 10 mg/mL)\*
- Collagen Buffer (5 mL)\*
- Ice bath
- 1x PBS
- 1M NaOH
- Container for mixing (15 mL Falcon tube or 5 mL Eppendorf tube)
- 3 mL syringes with Luer lock connections
- Female/female Luer lock adaptors\*
- Cells\* + culture medium\*
- Cartridges, 3 cc\*
- Sterile conical bioprinting nozzles\*
- Temperature-controlled printhead\* (optional)
- BIO X\*, BIO X6\* or INKREDIBLE-series\* 3D bioprinter
- Well plate or mold

\*The product can be purchased in the CELLINK shop at [www.cellink.com/shop/](http://www.cellink.com/shop/).

# Protocol for neutralization

During the whole procedure keep all materials on ice. Cool down the Coll 1 solution after every component is added.

## 1. Preparing Coll 1

### MATERIAL

Coll 1 Solution  
Collagen Buffer  
Ice bath

### DESCRIPTION

- Place the vial of Coll 1 Solution and the Collagen Buffer on ice to keep them cool.
- $C_S$  is the concentration of the original Coll 1 Solution (10 mg/mL).
- Record the desired final volume of the bioink ( $V_{INK}$ , mL).
- Record the desired final Coll 1 concentration after neutralization ( $C_F$ ).

Note:  $C_F$  and  $C_S$  cannot be the same, otherwise the solution would not be neutralized (refer to Table 1).

**Table 1.** Preparation of Coll 1 biomaterial with different concentration.

$V_{INK}$ , mL	$C_F$ , mg/mL	$C_S$ , mg/mL	$V_{Coll\ 1}$ , $\mu L$	$V_{CB}$ , $\mu L$	$V_{NaOH}$ , $\mu L$	$V_{PBS}$ , $\mu L$
1	8	10	800	123	20	57
	6		600	92	15	293
	4		400	62	10	528

## 2. Calculations for neutralization

### DESCRIPTION

- Volume of needed Coll 1 Solution:  $V_{Coll\ 1} (mL) = \frac{C_F \times V_{INK}}{C_S}$
- Volume of Collagen Buffer:  $V_{CB} (mL) = V_{Coll\ 1} \times 0.154$
- Volume of 1M NaOH:  $V_{NaOH} (mL) = V_{Coll\ 1} \times 0.025$
- Volume of 1x PBS to reach  $C_F$ :  $V_{PBS} (mL) = V_{INK} - V_{Coll\ 1} - V_{CB} - V_{NaOH}$

## 3. Neutralization

### MATERIAL

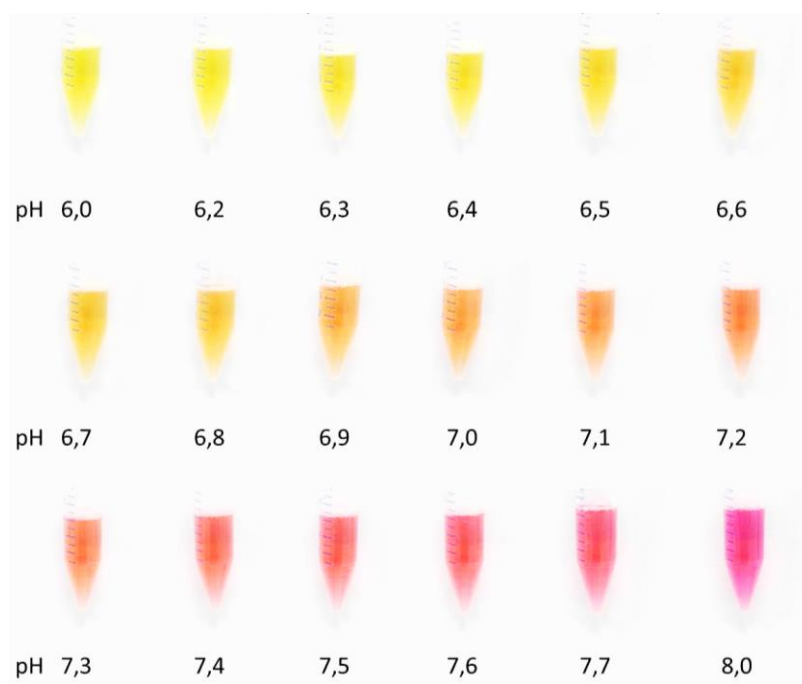
1x PBS  
1M NaOH  
Container for mixing  
Ice bath

## DESCRIPTION

- Mix  $V_{Coll\ 1}$  and  $V_{CB}$  in a sterile container with a suitable volume capacity by vortexing or pipetting up and down. Be extra careful with keeping the solution cool once the  $V_{CB}$  is added as Coll 1 may self-assemble if heated.
- Add  $V_{NaOH}$  to the mixing container.

Note: The natural material oxidation after the vial opening may affect the material pH. Therefore, if using previously opened vial, it is recommended to proceed by adding  $V_{NaOH}$  by smaller volume steps, until the color of the solution corresponds to a pH between 6.9-7.3 (refer to Figure 1). Note down the volume used. Cool on ice.

- Add  $V_{PBS}$  to the mixing container and homogenize by pipetting up and down or by vortexing.



**Figure 1.** Illustration of solution color correspondence to pH [1].

[1]<https://bit.ly/3ImRoCv>

# Protocol for bioprinting of neutralized Coll 1 solution

Make sure to follow the neutralization protocol above prior to following the bioprinting protocol. This bioprinting protocol works best using the BIO X/BIO X6 equipped with the Temperature-controlled printhead. If using the INKREDIBLE system, the bioprinting procedure should be performed fast to prevent the solution from warming and gelling in the cartridge during the experiment.

## 1. Prepare Coll 1 for mixing

### MATERIAL

Neutralized Coll 1 solution  
3 mL syringe with Luer lock connections

### DESCRIPTION

- Cool down the neutralized Coll 1 on ice for 10 min to make sure it remains in the liquid state.
- Transfer the solution into a 3 mL syringe using the following procedure: remove the syringe plunger → cap the syringe with a tip cap → pour the Coll 1 solution in the syringe → insert the plunger → flip the syringe → release the tip cap to evacuate the air.
- *If cells will be post-seeded on printed constructs, move directly to Step 3.*

## 2. Mixing Coll 1 with cells

### MATERIAL

Cell suspension in a syringe  
Cooled Coll 1 solution  
Female/female Luer lock adaptor

### DESCRIPTION

- Mix ten parts of Coll 1 solution with one part of cell suspension without introducing air bubbles to the mixture. For detailed instructions see the *Mixing cells with bioink Protocol*.
  - Attach the Coll 1 solution syringe to the syringe with cell suspension using a female/female Luer lock adaptor.
  - Carefully mix the solution with the cell suspension by gently pushing them back and forth between the syringes.

Note: Suggested cell suspension density is  $5 \times 10^6$  cells/mL to  $10 \times 10^6$  cells/mL.

Note: To avoid an air gap when mixing the solution and the cell suspension, carefully pre-fill the Luer lock adaptor with Coll 1 solution before attaching the syringe with the cell suspension.

## 3. Preparing for print

### MATERIAL

Temperature-controlled printhead (optional)  
Cartridge, 3 cc  
Coll 1 mixed with cells  
Conical bioprinting nozzles

## DESCRIPTION

- If using the BIO X/BIO X6, pre-cool the Temperature-controlled printhead to 5°C. If using the INKREDIBLE-series, cool down the cartridge on ice if needed.

Note: The bioprinting temperature can be increased to 10°C or 15°C, but this will reduce the time available for Coll 1 bioprinting process prior to its inadvertent self-assembly.

- Transfer the solution (with or without cells) to the cartridge, cap it with a bioprinting nozzle of choice and place it in the printhead.

# 4. Printing

## MATERIAL

BIO X, BIO X6 or INKREDIBLE series bioprinter

Well plate or mold

## DESCRIPTION

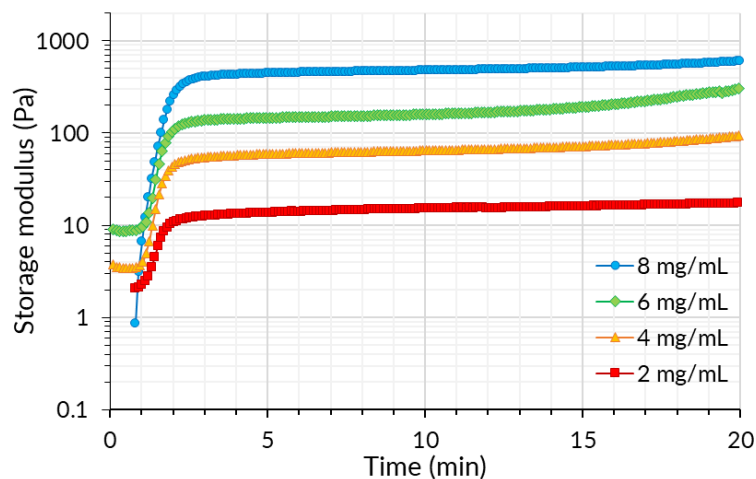
- Print droplets with the desired size in a mold or well plate.

# 5. Crosslinking

## DESCRIPTION

- Coll 1 can be crosslinked via thermal gelation.
  - Warm the bioprinted construct to 37°C until gelation occurs, approx. 10-15 min. The BIO X/BIO X6 heated print bed or incubation can be alternatively used. Refer to Figure 2 for thermal gelation behavior of Coll 1 with different final concentration.
  - *If cells were mixed with Coll 1 solution prior to bioprinting, move directly to Step 7.*

Note: The crosslinking time might be adjusted based on the construct thickness.



**Figure 2.** Thermal gelation of neutralized Coll 1 solution with different collagen concentrations ( $C_F$ ) indicated as storage moduli increase over time at 37°C.

# 6. Cell post-seeding

## MATERIAL

Cell suspension

#### DESCRIPTION

- Dispense the cell suspension in the middle of the printed hydrogel. Suggested cell suspension density:  $20 \times 10^3$  cells/cm<sup>2</sup> to  $50 \times 10^3$  cells/cm<sup>2</sup> (a highly concentrated cell suspension is suggested).

## 7. Incubation

#### MATERIAL

Cell culture medium

#### DESCRIPTION

- Add the desired medium to submerge the constructs and place in incubator.

Note: Ensure that the bioprinted constructs are crosslinked and do not dissolve in warm media.

- Incubate the constructs in cell culture medium in standard culture conditions (37°C, 5% CO<sub>2</sub> and 95% relative humidity) or according to your application.