This is a suggested procedure, please adjust according to your experimental needs. To maintain the sterility of the product, work under sterile conditions.

## Protocol aim

The aim of this protocol is to provide instructions for bioprinting of GeIXA bioink using the INKREDIBLE, INKREDIBLE+, BIO X or BIO X6, with and without cells. This document covers pre-print mixing with cells, 3D bioprinting and post-print processes such as ionic crosslinking and photocuring. This protocol was optimized for GeIXA with LAP $0.25 \%$ undiluted as well as a 10+1 cell suspension dilution. Changing the concentration of LAP or bioink to cell suspension ratio will change the photocrosslinking time. Refer to the Photocrosslinking Optimization Protocol to adjust and determine these numbers. This protocol was optimized using the Temperature-controlled printhead using the BIO X and BIO X6.

## Materials needed

- GeIXA bioink*
- Cells* + cell culture medium*
- 3 mL syringes with Luer lock connections
- Female/female Luer lock adaptor*
- CELLMIXER* (optional)
- UV shielding cartridges, 3 cc *
- Conical bioprinting nozzles, 22-27G recommended*
- Temperature-controlled printhead (optional)
- BIO X*, BIO X6* or INKREDIBLE series* 3D bioprinter
- Well plate or Petri dish*
- Crosslinking Agent* (included with the bioink purchase)

[^0]
## Protocol

This protocol works best using the BIO X or BIO X 6 with the cooled print bed at $15^{\circ} \mathrm{C}$ and the Temperaturecontrolled printhead at $24^{\circ} \mathrm{C}$. The GeIXA can also be extruded using the pneumatic printheads or the INKREDIBLE series, but with decreased shape fidelity if ambient temperature exceeds $25^{\circ} \mathrm{C}$ and the printhead heats up. If using the INKREDIBLE series, the printing substrates such as Petri dishes or well plates should be placed on ice or another cooled surface to thermally gel the construct after printing prior to photocrosslinking. First time users of GeIMA based bioinks are recommended to optimize the printing conditions without cells before proceeding to bioprint with cells. Perform the desired dilution using medium or PBS.

## 1. Preparing the bioink

## MATERIAL

## GeIXA

3 mL syringe with Luer lock connections
Female/female Luer lock adaptor
Pipette tip or spatula

## DESCRIPTION

- Heat up the GeIXA cartridge at $37^{\circ} \mathrm{C}$ until it becomes liquid. The heating of the GeIXA can be performed in a printhead or incubator and usually requires $30-60 \mathrm{~min}$ if the bioink is taken directly from the fridge. Note: Keep the cartridge horizontal when tip and end cap are removed. This is to prevent air from entering the cartridge or bioink from dripping out.
- To make sure that the bioink is homogeneous after heating, connect the cartridge to a 3 mL syringe using a Luer lock connector and remove the end cap. Push approximately half of the bioink from the cartridge into the syringe by gently pushing the cartridge piston using a pipette tip or small spatula while simultaneously pulling the syringe plunger. To remove any air bubble derived from the dead volume in the syringe, separate the syringe and cartridge maintaining the Luer lock on the syringe. Hold the syringe with the tip facing upwards and gently tap the syringe to move air bubbles towards the tip. Carefully extrude air and pre-fill the Luer lock adaptor with GeIXA before re-attaching the cartridge. Gently mix the bioink back and forth between the cartridge and syringe to homogenize the bioink. If not using the entire 3 mL of the bioink in the cartridge, keep the rest of the bioink in the optimal storage conditions. Prolonged and repeated heating could negatively affect the photoinitiator stability.
Note: If there are bubbles in the bioink, make a quick centrifugation for 30 s at $600 \times \mathrm{g}$.
- If not printing with cells move directly to Step 3.


## 2. Mixing the bioink with cells

## MATERIAL

3 mL syringes with Luer lock connections
Female/female Luer lock adaptor
Pre-warmed GeIXA
Cell suspension
UV shielding cartridge, 3cc
CELLMIXER (optional)

## DESCRIPTION

- At this point, mix ten parts of bioink with one part of cell suspension, taking care not to introduce air bubbles to the mixture. For detailed instructions see the Mixing cells with bioink Protocol.
- If preparing for quantities $<2 \mathrm{~mL}$ of GelXA, it is recommended to connect two 3 mL Luer lock syringes, one with the bioink and the other with the cell suspension and gently mix back and forth between the syringes until homogeneous. Transfer the mixture to an empty 3cc cartridge by connecting the syringe to the cartridge using the Luer lock adaptor. Cap the cartridge with a tip cap.
- If using larger quantities, the CELLMIXER can be used:
- Transfer the cell suspension to the 1 mL cell syringe (PART 1) using a female/female Luer lock adaptor.
- Transfer GeIXA to the 12 mL syringe (PART 2) using a female/female Luer lock adaptor.
- Clip both syringes to the dispensing unit (PART 3).
- Connect the two syringes to the mixing unit (PART 4), then connect the Empty cartridge (PART 5) to the Mixing unit's other side.
- Apply gentle pressure onto the dispensing unit to mix the content of both syringes into the empty cartridge. Cap the cartridge with a tip cap.
Note: To avoid introducing air when connecting the syringes, carefully pre-fill the Luer lock adaptor with GeIXA before attaching it to the syringe with the cell suspension.


## 3. Preparation for printing

## MATERIAL

GeIXA mixed with cells (if applicable) in UV shielding cartridge
Temperature-controlled printhead (optional)
Conical bioprinting nozzles, 22-27G

## DESCRIPTION

- If the cartridge is just taken from the heat or if the cartridge still feels warm after mixing in the cells, place it in the pre-heated Temperature-controlled printhead at $24^{\circ} \mathrm{C}$ for 10 minutes. If not using the Temperature-controlled printhead, place the cartridge on counter for 5-10 minutes to reach approximately $24^{\circ} \mathrm{C}$.
- If the cartridge has cooled down below $23^{\circ} \mathrm{C}$, re-heat the cartridge at $37^{\circ} \mathrm{C}$ for 5 min to reset, then restart equilibration in printhead.
- Cap the cartridge with a bioprinting nozzle and place the GeIXA in the printhead. Connect the cartridge to the air pressure adapter. If using the $\mathrm{BIO} X$ or $\mathrm{BIO} X 6$, pre-cool the print bed to $15^{\circ} \mathrm{C}$.
Note: When printing with GeIXA, the recommended printhead temperature for the highest printing fidelity is $24^{\circ} \mathrm{C}$, though the bioink can be dispensed up to $32^{\circ} \mathrm{C}$. Below $23^{\circ} \mathrm{C}$ the bioink can become too viscous resulting in chunky filaments and too high extrusion pressures needed.
Note: Be careful not to touch the printhead with the nozzle tip and if using very liquid materials, make sure that the bioink does not drip through the nozzle especially when attaching the air adapter. Alternatively, the cartridge can be placed in the printhead with the tip cap on and when in place switched to a nozzle.
Note: Test the flow of the bioink after the calibration is performed and start with a low pressure and increase stepwise.


## 4. Printing

## MATERIAL

BIO X, BIO X6 or INKREDIBLE series bioprinter
Well plate or Petri dish

## DESCRIPTION

- Calibrate the nozzle to the well plate or Petri dish surface. Test the flow of the bioink first after calibration and start with a low pressure and increase stepwise. Bioprint structures into the well plate or Petri dish. If printability is not as desired, adjust the pressure up/down by 1 kPa to extrude more/less material.
Example: If printing continuous filaments with a Temperature-controlled printhead set to $24^{\circ} \mathrm{C}$, a 25 G nozzle, a printing speed of $5 \mathrm{~mm} / \mathrm{s}$ and with 300 ms pre-flow delay, the suggested pressure range is between 27-37 kPa without cell suspension dilution and $22-32 \mathrm{kPa}$ if diluted with cell suspension.
- If proper viscosity and printability is not achieved by extending temperature equilibration time or tuning pressure:
- Too low viscosity (wide filaments despite using low pressure): decrease the printhead temperature $0.5-1^{\circ} \mathrm{C}$ to increase the viscosity and equilibrate an additional couple of minutes.
- Too high viscosity (chunky filaments and high pressure required): increase the printhead temperature $0.5-1^{\circ} \mathrm{C}$ to decrease the viscosity and equilibrate an additional couple of minutes. If unsuccessful, the bioink might have over-gelled. In this case, re-heat the cartridge at $37^{\circ} \mathrm{C}$ for 5 min to reset and choose a $1^{\circ} \mathrm{C}$ higher printhead temperature than before. Let the bioink reach the new temperature before starting to print again.
- During print sessions longer than 20 min at $24^{\circ} \mathrm{C}$ the bioink can become too viscous due to continued gelling resulting in chunky filaments and too high extrusion pressures needed. To avoid this, a $0.5^{\circ} \mathrm{C}$ increase in printhead temperature after 20 min printing can extend the time the bioink remains at good printability.
- If the regular pneumatic printheads are used for a long period, they might heat up above the desired printing temperature. Then the bioink also heats up which decreases its viscosity, observed as extrusion of very thick filaments even at low pressures. Remove the cartridge from the printhead and allow to cool down to $24^{\circ} \mathrm{C}$. In addition, remove the printheads from the BIO X/BIO X6 to let them cool down before continuing to print.
Note: If waiting too long between extrusions the bioink can dry in the nozzle causing it to clog. If this occurs, replace with new nozzle.


## 5. crosslinking

## MATERIAL

Crosslinking Agent
AND/OR
405 or 365 nm LED modules for photocuring
Cell culture medium

## DESCRIPTION

- GeIXA can be photocrosslinked using the 405 or 365 nm LED modules or ionically crosslinked using the $\mathrm{CaCl}_{2}$-containing Crosslinking Agent. If using both, begin with photocrosslinking. It is recommended to use either photocrosslinking alone for softer constructs or both crosslinking methods which will generate more robust constructs. Optimize the crosslinking according to application.
Note: It is recommended to use the 405 nm LED module instead of 365 nm if possible. Overexposure might damage the cells.
- Photocrosslinking: See Table 1 below for recommended photocrosslinking times. Ensure that the bioprinted GeIXA construct is thermally gelled after printing by cooling the print bed (if using the BIO X or BIO X6) or placing the printing substrates with the construct on ice for 10 seconds (if using the INKREDIBLE series). If photocrosslinking during bioprinting, set the crosslinking parameters appropriately in the G-code for the INKREDIBLE series or the printhead setup page for the BIO X or BIO X6.
Note: To verify that the photocrosslinking is sufficient, add $37^{\circ} \mathrm{C}$ media to one printed well and observe that it doesn't dissolve.
- Ionic crosslinking: Submerge the cell-laden constructs in the Crosslinking Agent for 30 seconds to 5 minutes depending on construct size, infill density and desired construct stiffness. Remove Crosslinking Agent and rinse constructs with basal culture media once.
Note: 30 seconds is recommended for $10 \mu \mathrm{~L}$ droplets while 10 minutes might be required for dense $1 \mathrm{~cm}^{3}$ blocks. In addition, optimize the crosslinking depending on the cell type.

Table 1. Recommended time of the construct photocrosslinking**. Distance from each light module to construct was set to 5 cm using the BIO X or BIO X6 photocuring modules. If using the INKREDIBLE series photocuring modules, the time required can possibly be decreased. For crosslinking with other parameters, see Photocrosslinking Optimization Protocol. This table was generated using GelXA with mesenchymal stem cells. Don't exceed 120 seconds of exposure time when printing with cells. To achieve the best structural integrity when printing thicker constructs, it is recommended to apply 3 or 5 seconds photocrosslinking with 365 or 405 nm light respectively, every fourth layer. If the bioink is used above $25^{\circ} \mathrm{C}$, the best results can be achieved when photocrosslinking every second layer.

|  | 365 nm, LAP 0.25\% | 405 nm, LAP 0.25\% |
| :---: | :---: | :---: |
| $\mathbf{1 m m}$ construct thickness | 5 seconds | 10 seconds |
| 3 mm construct thickness | 15 seconds | 30 seconds |

${ }^{* *}$ This is only a recommended reference of crosslinking times to start with. The actual time needed for crosslinking will vary depending on the size and temperature of the constructs as well as the intensity of the photocuring module and the distance to the construct.

## 6. Incubation

## MATERIAL

Cell culture medium

## DESCRIPTION

- After crosslinking, add the desired medium to the constructs and place them in an incubator.
- Incubate the constructs in cell culture medium in standard culture conditions $\left(37^{\circ} \mathrm{C}, 5 \% \mathrm{CO}_{2}\right.$ and $95 \%$ relative humidity) or according to application. Replace medium regularly.


[^0]:    *The product can be purchased in the CELLINK shop at www.cellink.com/shop.

